

101.215-12 – including *Taq* pol., IFU-01
101.215-12u – without *Taq* pol., IFU-02

Visit <https://labproducts.caredx.com> for
“Instructions for Use” (IFU)

Lot No.: **7H9**

Lot-specific Information
Olerup SSP[®] DQB1*04

Product number:	101.215-12 – including <i>Taq</i> polymerase 101.215-12u – without <i>Taq</i> polymerase
Lot number:	7H9
Expiry date:	2023-05-01
Number of tests:	12
Number of wells per test:	20+1
Storage - pre-aliquoted primers:	dark at -20°C
- PCR Master Mix:	-20°C
- Adhesive PCR seals	RT
- Product Insert	RT

This Product Description is only valid for Lot No. 7H9.

Complete product documentation consists of generic Instructions for Use (IFU), lot specific Product Insert, Worksheet and Certificate.

**CHANGES COMPARED TO THE PREVIOUS OLERUP SSP[®]
DQB1*04 Lot (5G5)**

The DQB1*04 kit is updated for new alleles to enable separation of:

- Null and Alternatively expressed alleles.

The product documentation has been updated for new alleles of IMGT 3.32.0.

The DQB1*04 specificity and interpretation tables have been updated with the DQB1 alleles described since the previous *Olerup SSP[®] DQB1*04 lot (Lot No. 5G5)* was made.

The DQB1*04 primer set is unchanged compared to the previous *Olerup SSP[®] DQB1*04 Lot (Lot No. 5G5)*.

Changes in revision R01 compared to R00:

1. The expiration date has been altered due to shelf-life extension.

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Well **21** contains Negative Control primer pairs, that will amplify a majority of the *Olerup* SSP[®] HLA Class I, DRB, DQB1, DPB1 and DQA1 amplicons as well as all the amplicons generated by the control primer pairs matching the human growth hormone gene.

HLA-specific PCR product sizes range from 75 to 200 base pairs.
The PCR product generated by the positive control primer pair is 430 base pairs.

Length of PCR product	105	200	105	80	75	80	85
5'-primer¹	164	340	440	45	45	43	36
	5'-CAC ^{3'}	5'-Agg ^{3'}	5'-TTA ^{3'}	5'-Tgg ^{3'}	5'-Tgg ^{3'}	5'-Tgg ^{3'}	5'-TAC ^{3'}
							36
							5'-TAT ^{3'}
3'-primer²	231	2nd I	507	59	58	57	47
	5'-TgC ^{3'}	5'-AAA ^{3'}	5'-TTg ^{3'}	5'-CTC ^{3'}	5'-ggC ^{3'}	5'-CTC ^{3'}	5'-ACA ^{3'}
							48
							5'-gCA ^{3'}
							48
							5'-gCC ^{3'}
							52
							5'-TgT ^{3'}
A*	+	+	+				
B*	+	+	+				
C*	+	+	+				
DRB1				+	+		
DRB3				+	+		
DRB5				+			
DQB1					+		
DPB1						+	
DQA1							+

¹The nucleotide position for HLA class I genes and the codon for HLA class II genes, in the 2nd or 3rd exon, matching the specificity-determining 3'-end of the primer is given. Nucleotide and codon numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

²The nucleotide position for HLA class I genes and the codon for HLA class II genes, in the 2nd or 3rd exon or the 2nd intron, matching the specificity-determining 3'-end of the primer is given in the anti-sense direction. Nucleotide and codon numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

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PRODUCT DESCRIPTION

DQB1*04 SSP subtyping

CONTENT

The primer set contains 5'- and 3'-primers for identifying the DQB1*04:01 to DQB1*04:42 alleles.

Please note that DQB1 amplifications usually are somewhat less pronounced than e.g. DRB and DQA1 amplifications even when using the same DNA preparation and exactly the same experimental procedures.

PLATE LAYOUT

Each test consists of 21 PCR reactions in a 24 well PCR plate. Wells 22 to 24 are empty.

1	2	3	4	5	6	7	8
9	10	11	12	13	14	15	16
17	18	19	20	NC	empty	empty	empty

The 24 well PCR plate is marked with ‘DQB1*04’ in silver/gray ink.

Well No. 1 is marked with the Lot No. ‘7H9’.

Wells 1 to 20 – DQB1*04 high resolution primers.

Well 21 – Negative Control (NC).

A faint row of numbers is seen between wells 1 and 2 or wells 7 and 8 of the PCR trays. These stem from the manufacture of the trays, and should be disregarded. The PCR plates are covered with a PCR-compatible foil.

Please note: When removing each 24 well PCR plate, make sure that the remaining plates stay covered. Use a scalpel or a similar instrument to carefully cut the foil between the plates.

INTERPRETATION

Due to the sharing of sequence motifs between DQB1 alleles, non-DQB1*04 alleles will be amplified by some primer mixes.

The interpretation of DQB1*04 subtypings is not influenced by the DQB2 and DQB3 genes.

For further details see Specificity Table.

UNIQUELY IDENTIFIED ALLELES

All the DQB1*04 alleles, i.e. **DQB1*04:01 to DQB1*04:42**, recognized by the HLA Nomenclature Committee in April 2018^{1,2} will be amplified by the primers in the DQB1*04 subtyping kit.

The DQB1*04 kit enables separation of the confirmed DQB1*04 alleles as listed in the IMGT/HLA database 3.25.0. An HLA allele is listed as confirmed by IMGT/HLA

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if it has been sequenced by more than a single laboratory or from multiple sources. Current allele confirmation status for DQB1*04 alleles is listed below.

The DQB1*04 kit also enables identification of null and alternatively expressed alleles.

¹HLA-DQB1 alleles listed on the IMGT/HLA web page 2018-April-16, release 3.32.0, www.ebi.ac.uk/imgt/hla.

²Alleles that have been deleted from or renamed in the official WHO HLA Nomenclature up to and including the last IMGT/HLA database release can be retrieved from web page <http://hla.alleles.org/alleles/deleted.html>.

ALLELE CONFIRMATION STATUS

Allele	Status ¹	Allele	Status ¹	Allele	Status ¹
DQB1*04:01:01	Confirmed	DQB1*04:08	Unconfirmed	DQB1*04:28	Unconfirmed
DQB1*04:01:02	Unconfirmed	DQB1*04:09	Unconfirmed	DQB1*04:29	Unconfirmed
DQB1*04:01:03	Unconfirmed	DQB1*04:10	Unconfirmed	DQB1*04:30	Unconfirmed
DQB1*04:01:04	Unconfirmed	DQB1*04:11	Confirmed	DQB1*04:31	Unconfirmed
DQB1*04:02:01	Confirmed	DQB1*04:12	Confirmed	DQB1*04:32	Unconfirmed
DQB1*04:02:02	Unconfirmed	DQB1*04:13	Unconfirmed	DQB1*04:33	Confirmed
DQB1*04:02:03	Confirmed	DQB1*04:14	Unconfirmed	DQB1*04:34	Unconfirmed
DQB1*04:02:04	Confirmed	DQB1*04:15	Unconfirmed	DQB1*04:35	Unconfirmed
DQB1*04:02:05	Unconfirmed	DQB1*04:16	Unconfirmed	DQB1*04:36N	Unconfirmed
DQB1*04:02:06	Unconfirmed	DQB1*04:17	Unconfirmed		
DQB1*04:02:07	Confirmed	DQB1*04:18	Confirmed		
DQB1*04:02:08	Unconfirmed	DQB1*04:19	Unconfirmed		
DQB1*04:02:09	Confirmed	DQB1*04:20	Unconfirmed		
DQB1*04:02:10	Unconfirmed	DQB1*04:21	Unconfirmed		
DQB1*04:03:01	Unconfirmed	DQB1*04:22	Unconfirmed		
DQB1*04:03:02	Unconfirmed	DQB1*04:23	Unconfirmed		
DQB1*04:04	Unconfirmed	DQB1*04:24	Unconfirmed		
DQB1*04:05	Unconfirmed	DQB1*04:25N	Unconfirmed		
DQB1*04:06	Unconfirmed	DQB1*04:26	Unconfirmed		
DQB1*04:07	Unconfirmed	DQB1*04:27	Confirmed		

¹Allele status “confirmed” or “unconfirmed” as listed on the IMGT/HLA web page 2016-July-14, release 3.25.0, www.ebi.ac.uk/imgt/hla.

RESOLUTION IN HOMO- AND HETEROZYGOTES

Results file with resolution in DQB1*04 homo- and heterozygotes is available upon request.

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Lot-specific Information
SPECIFICITY TABLE

DQB1*04 SSP subtyping

Specificities and sizes of the PCR products of the 20+1 primer mixes used for DQB1*04 SSP subtyping

Primer Mix	Size of spec. PCR product ¹	Size of control band ²	Amplified DQB1*04 alleles ³	Amplified non-DQB1*04 alleles
1^{5,6}	160 bp 205 bp	515 bp	*04:01:01:01-04:01:02, 04:01:04-04:02:01:01, 04:02:01:04-04:02:07, 04:02:09-04:03:02, 04:04- 04:29, 04:31-04:42 *04:01:01:01-04:01:05, 04:05-04:08, 04:14-04:17, 04:38, 04:42	*03:132
2^{5,6}	205 bp	515 bp	*04:01:01:01-04:01:05, 04:05-04:08, 04:14-04:17, 04:38, 04:42	
3	180 bp	430 bp	*04:02:01:01, 04:02:01:04- 04:03:01, 04:04, 04:09- 04:13, 04:18-04:19, 04:21, 04:22 ^w , 04:23, 04:24 ^w , 04:25N-04:32, 04:34-04:37, 04:39-04:41N	
4	195 bp	430 bp	*04:03:01-04:03:03	*03:06, 03:25:01
5⁴	110 bp 245 bp	430 bp	*04:06, 04:12 *04:04-04:05	*03:06, 03:25:01
6⁴	95 bp 210 bp	430 bp	*04:16 *04:20	
7⁷	245 bp	430 bp	*04:02:08	*03:30, 03:72, 03:100, 03:215, 06:02:01:01-06:02:28, 06:10, 06:13:01-06:16, 06:19:01- 06:20, 06:22:01-06:24, 06:29, 06:33, 06:37, 06:46-06:51:02, 06:68-06:84, 06:95-06:97, 06:106-06:107, 06:109, 06:111-06:117, 06:119, 06:122-06:123, 06:125-06:127, 06:130, 06:136, 06:138- 06:139, 06:146:01-06:147, 06:150-06:152, 06:156, 06:159, 06:161-06:163, 06:166, 06:173-06:175, 06:178-06:179N, 06:182- 06:183, 06:188, 06:192, 06:197-06:198, 06:200, 06:206:01-06:206:02, 06:208, 06:211, 06:213, 06:215- 06:216N, 06:219,

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				06:224-06:228, 06:232, 06:235-06:237, 06:240, 06:242, 06:249
8⁴	95 bp	430 bp	*04:08	*03:06 [?] -03:07 [?] , 03:11 [?] , 03:14:01 [?] -03:15 [?] , 03:17:01 [?] - 03:18 [?] , 03:19:01-03:19:02, 03:20 [?] , 03:23:01 [?] , 03:25:02 [?] - 03:26 [?] , 03:37 [?] , 03:40 [?] , 03:48 [?] , 03:52 [?] -03:71 [?] , 03:74 [?] -03:78 [?] , 03:81 [?] -03:82 [?] , 03:101 [?] - 03:104 [?] , 03:106 [?] -03:112 [?] , 03:118N [?] , 03:120 [?] -03:131 [?] , 03:133 [?] -03:135 [?] , 03:137 [?] , 03:140 [?] -03:149 [?] , 03:150, 03:151 [?] -03:163 [?] , 03:165 [?] - 03:167 [?] , 03:170 [?] -03:179 [?] , 03:183 [?] , 03:185 [?] , 03:187 [?] - 03:189 [?] , 03:192 [?] -03:194 [?] , 03:201 [?] -03:205 [?] , 03:207 [?] - 03:210 [?] , 03:212 [?] -03:221 [?] , 03:223 [?] -03:230 [?] , 03:232 [?] - 03:238 [?] , 03:240 [?] , 03:244 [?] , 03:255 [?] -03:262 [?] , 03:263, 03:267 [?] -03:275 [?] , 03:278 [?]
9	140 bp	430 bp	*04:09, 04:14	
10	145 bp	430 bp	*04:10	*05:01:19 ^w , 06:03:03 ^w
11⁴	120 bp 160 bp	430 bp	*04:11, 04:15 *04:23	*03:22, 03:96, 03:251, 05:103 ^w , 06:04:01 ^w -06:05:02 ^w , 06:06 [?] , 06:07:01 ^w -06:07:02 ^w , 06:08:02 [?] -06:08:03 [?] , 06:09:01:01 ^w -06:09:07 ^w , 06:11:02 [?] -06:11:03 [?] , 06:12 ^w , 06:13:02 [?] , 06:15:01 [?] - 06:15:02 [?] , 06:17 [?] -06:18:01 [?] , 06:18:02 ^w , 06:19:01 [?] , 06:21 ^w , 06:22:01 [?] -06:25 [?] , 06:27:01 [?] - 06:32:02 [?] , 06:34 ^w , 06:35 [?] , 06:36 ^w , 06:38 ^w , 06:40 [?] , 06:41 ^w - 06:42 ^w , 06:45 [?] , 06:50 [?] , 06:51:02 [?] , 06:52 ^w , 06:53:01 [?] - 06:54N [?] , 06:56 [?] -06:57 [?] , 06:58 ^w , 06:59 [?] -06:61 [?] , 06:63 [?] - 06:67 [?] , 06:69:01 [?] -06:76 [?] , 06:79:02 [?] -06:83 [?] , 06:84 ^w , 06:85 [?] , 06:89 [?] , 06:91 [?] -06:97 [?] , 06:118:01 ^w , 06:118:02 [?] - 06:124 [?] , 06:126 [?] , 06:128 [?] , 06:129 ^w , 06:131 [?] -06:147 [?] , 06:149 [?] -06:184 [?] , 06:186 ^w , 06:189 ^w , 06:190:01 [?] -06:199 [?] , 06:201 [?] -06:204 [?] , 06:206:01 [?] ,

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				06:206:02 ^w , 06:207 [?] -06:208 [?] , 06:210 [?] -06:215 [?] , 06:217 ^w , 06:229 [?] -06:236 [?] , 06:238 [?] , 06:241 [?] , 06:244 [?] , 06:246 [?] - 06:247 [?] , 06:249 [?] , 06:252N [?]
12	160 bp 230 bp	430 bp	*04:07 *04:18	
13	185 bp	430 bp	*04:13	
14⁶	160 bp	430 bp	*04:17	
15	150 bp	430 bp	*04:23, 04:25N	*03:96, 03:251
16	200 bp	430 bp	*04:27	
17	195 bp	430 bp	*04:33	
18	245 bp	430 bp	*04:36N	
19	225 bp	430 bp	*04:01:01:01-04:02:01:01, 04:02:01:04-04:03:03, 04:06-04:21, 04:22 ^w , 04:23, 04:24 ^w , 04:25N-04:32, 04:34-04:37, 04:38 ^w , 04:39- 04:42	
20⁴	105 bp	430 bp	*04:41N	
21⁸	-	-	Negative Control	

¹Alleles are assigned by the presence of specific PCR product(s). However, the sizes of the specific PCR products may be helpful in the interpretation of DQB1*04 SSP typings.

When the primers in a primer mix can give rise to HLA-specific PCR products of more than one length this is indicated if the size difference is more than 20 base pairs. Size differences of 20 base pairs or less are not given. For high resolution SSP kits, the alleles listed are specified according to amplicon length.

Nonspecific amplifications, i.e. a ladder or a smear of bands, may sometimes be seen. GC-rich primers have a higher tendency of giving rise to nonspecific amplifications than other primers.

PCR fragments longer than the control bands may sometimes be observed. Such bands should be disregarded and do not influence the interpretation of the SSP typings.

PCR fragments migrating faster than the control bands, but slower than a 400 bp fragment may be seen in some gel read-outs. Such bands can be disregarded and do not influence the interpretation of the SSP typings.

Some primers may give rise to primer oligomer artifacts. Sometimes this phenomenon is an inherent feature of the primer pair(s) of a primer mix. More often it is due to other factors such as too low amount of DNA in the PCR reactions, taking too long time in setting up the PCR reactions, working at elevated room temperature or using thermal cyclers that are not pre-heated.

²The internal positive control primer pairs amplify segments of the human growth hormone gene. The internal positive control bands are 430 or 515 base pairs respectively, well distribution as outlined in the table. Well number 1 contains the longer, 515 bp, internal positive control band. The well distribution of the internal controls can help in orientation of the kit on gel photo, as well as allow for kit identification. In the presence of a specific amplification the intensity of the control band often decreases.

³For several DQB1 alleles 1st and/or 3rd exon(s) and beyond, as well as intron nucleotide sequences, are not available. In these instances it is not known whether some of the primers of the SSP sets are completely matched with the target sequences or not. Assumption is made that unknown sequences in these regions are conserved within allelic groups.

⁴HLA-specific PCR products shorter than 125 base pairs have a lower intensity and are less sharp than longer PCR products.

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⁵Primer mixes 1 and 2 may give rise to a lower yield of HLA-specific PCR product than the other DQB1*04 primer mixes.

⁶Primer mixes 1, 2 and 14 may have tendencies of unspecific amplifications.

⁷In primer mix 7 the positive control band may be weaker than for other DQB1*04 primer mixes.

⁸Primer mix 21 contains a negative control, which will amplify the majority of the HLA amplicons as well as the amplicons generated by the control primer pairs matching the human growth hormone gene. HLA-specific PCR product sizes range from 75 to 200 base pairs and the PCR product generated by the HGH positive control primer pair is 430 base pairs.

Abbreviations

'w', might be weakly amplified.

'?', nucleotide sequence information not available for the primer matching sequence.

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PRIMER SPECIFICATION

Well No.	1	2	3	4	5	6	7	8	9	10	11	12
Length of spec. PCR product	160 205	205	180	195	110 245	95 210	245	95	140	145	120 160	160 230
Length of int. pos. control ¹	515	515	430	430	430	430	430	430	430	430	430	430
5'-primer(s) ²	23(164) 5'-gCT 3'	23(164) 5'-gCT 3'	23(164) 5'-gCg 3'	26(173) 5'-TCT 3'	9(122) 5'-gTA 3'	22(160) 5'-CCA 3'	9(122) 5'-gTT 3'	167(596) 5'-gCA 3'	44(226) 5'-ACA 3'	23(164) 5'-gCg 3'	49(241) 5'-ggA 3'	15(140) 5'-gTA 3'
	38(210) 5'-gCg 3'		23(164) 5'-gCg 3'		55(259) 5'-ggT 3'	59(272) 5'-CgT 3'			45(230) 5'-ggA 3'		52(251) 5'-gCT 3'	145(529) 5'-CCg 3'
											130(485) 5'-CCA 3'	
3'-primer(s) ³	77(327) 5'-ACg 3'	77(327) 5'-ACg 3'	70(304) 5'-CTC 3'	77(327) 5'-ACg 3'	77(327) 5'-ACg 3'	77(327) 5'-ACg 3'	77(326) 5'-CCg 3'	185(650) 5'-CgA 3'	77(327) 5'-ACg 3'	57(267) 5'-gCA 3'	77(327) 5'-ACg 3'	77(327) 5'-ACg 3'
											170(604) 5'-gAC 3'	185(650) 5'-CgA 3'
Well No.	1	2	3	4	5	6	7	8	9	10	11	12

Well No.	13	14	15	16	17	18	19	20
Length of spec. PCR product	185	160	150	200	195	245	225	105
Length of int. pos. control ¹	430	430	430	430	430	430	430	430
5'-primer(s) ²	140(514) 5'-CAA 3'	146(533) 5'-CCT 3'	26(173) 5'-ggg 3'	24(166) 5'-gCT 3'	23(164) 5'-gCg 3'	26(173) 5'-ggg 3'	9(122) 5'-gTT 3'	102(400) 5'-TCT 3'
			130(485) 5'-CCA 3'					
3'-primer(s) ³	187(656) 5'-ACA 3'	185(650) 5'-CgA 3'	59(271) 5'-CTA 3'	77(327) 5'-ACg 3'	74(316) 5'-CgC 3'	94(376) 5'-TCA 3'	70(304) 5'-CTC 3'	123(465) 5'-ggC 3'
			185(650) 5'-CgA 3'					
Well No.	13	14	15	16	17	18	19	20

¹The internal positive control primer pairs amplify segments of the human growth hormone gene. The internal positive control bands are 430 or 515 base pairs respectively, well distribution as outlined in the table. Well number 1 contains the longer, 515 bp, internal positive control band. The well distribution of the internal controls can help in orientation of the kit on gel photo, as well as allow for kit identification. In the presence of a specific amplification the intensity of the control band often decreases.

²The nucleotide position matching the specificity-determining 3'-end of the primer is given. Nucleotide numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

³The nucleotide position matching the specificity-determining 3'-end of the primer is given in the anti-sense direction. Nucleotide numbering as on the www.ebi.ac.uk/imgt/hla web site. The sequence of the 3 terminal nucleotides of the primer is given.

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CELL LINE VALIDATION SHEET																				
DQB1*04 SSP subtyping kit ²																				
				Well																
				1	2	3	4	5	6	7	8	9	10	11	12	13	14	15	16	
				201550701	201550702	201550703	201432104	201432105	201550706	201550707	201432108	201432109	201432110	201893611	201550712	201432113	201432114	201893615	201673716	
	IHC cell line ¹		DQB1	Production No.																
1	9001	SA	*05:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
2	9280	LK707	*06:01	*02:02	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
3	9011	E4181324	*06:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
4	9275	GU373	*02:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
5	9009	KAS011	*05:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
6	9353	SM	*03:02	*06:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
7	9020	QBL	*02:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
8	9025	DEU	*03:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
9	9026	YAR	*03:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
10	9107	LKT3	*04:01		+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	
11	9051	PITOUT	*02:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
12	9052	DBB	*03:03		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
13	9004	JESTHOM	*05:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
14	9071	OLGA	*04:02		+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	
15	9075	DKB	*03:03		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
16	9037	SWEIG007	*03:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
17	9282	CTM3953540	*02:01	*06:03	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
18	9257	32367	*06:02	*02:02	-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	
19	9038	BM16	*03:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
20	9059	SLE005	*06:04		-	-	-	-	-	-	-	-	-	W	-	-	-	-	-	
21	9064	AMALA	*03:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
22	9056	KOSE	*05:03	*06:04	-	-	-	-	-	-	-	-	-	W	-	-	-	-	-	
23	9124	IHL	*05:03	*06:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
24	9035	JBUSH	*03:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
25	9049	IBW9	*02:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
26	9285	WT49	*02:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
27	9191	CH1007	*04:01	*05:01	+	+	-	-	-	-	-	-	-	-	-	-	-	-	-	
28	9320	BEL5GB	*02:02	*03:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
29	9050	MOU	*02:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
30	9021	RSH	*04:02		+	-	+	-	-	-	-	-	-	-	-	-	-	-	-	
31	9019	DUCAF	*02:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
32	9297	HAG	*03:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
33	9098	MT14B	*03:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
34	9104	DHIF	*03:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
35	9302	SSTO	*03:05		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
36	9024	KT17	*03:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
37	9065	HHKB	*06:03		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
38	9099	LZL	*03:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
39	9315	CML	*02:01	*03:01	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
40	9134	WHONP199	*02:02	*03:03	-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
41	9055	H0301	*06:09		-	-	-	-	-	-	-	-	-	W	-	-	-	-	-	
42	9066	TAB089	*06:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
43	9076	T7526	*03:03		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
44	9057	TEM	*05:03		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
45	9239	SHJO	*02:02		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
46	9013	SCHU	*06:02		-	-	-	-	-	+	-	-	-	-	-	-	-	-	-	
47	9045	TUBO	*03:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	
48	9303	TER-ND	*05:01		-	-	-	-	-	-	-	-	-	-	-	-	-	-	-	



101.215-12 – including *Taq* pol., IFU-01
 101.215-12u – without *Taq* pol., IFU-02

Visit <https://labproducts.caredx.com> for
 “Instructions for Use” (IFU)

Lot No.: **7H9**

Lot-specific Information

CELL LINE VALIDATION SHEET								
DQB1*04 SSP subtyping kit ²								
				Well				
				17	18	19	20	
				201673717	201673718	201893619	201893620	
				Production No.				
IHCW cell line ¹			DQB1					
1	9001	SA	*05:01		-	-	-	-
2	9280	LK707	*06:01	*02:02	-	-	-	-
3	9011	E4181324	*06:01		-	-	-	-
4	9275	GU373	*02:01		-	-	-	-
5	9009	KAS011	*05:02		-	-	-	-
6	9353	SM	*03:02	*06:01	-	-	-	-
7	9020	QBL	*02:01		-	-	-	-
8	9025	DEU	*03:01		-	-	-	-
9	9026	YAR	*03:02		-	-	-	-
10	9107	LKT3	*04:01		-	-	+	-
11	9051	PITOUT	*02:02		-	-	-	-
12	9052	DBB	*03:03		-	-	-	-
13	9004	JESTHOM	*05:01		-	-	-	-
14	9071	OLGA	*04:02		-	-	+	-
15	9075	DKB	*03:03		-	-	-	-
16	9037	SWEIG007	*03:01		-	-	-	-
17	9282	CTM3953540	*02:01	*06:03	-	-	-	-
18	9257	32367	*06:02	*02:02	-	-	-	-
19	9038	BM16	*03:01		-	-	-	-
20	9059	SLE005	*06:04		-	-	-	-
21	9064	AMALA	*03:01		-	-	-	-
22	9056	KOSE	*05:03	*06:04	-	-	-	-
23	9124	IHL	*05:03	*06:01	-	-	-	-
24	9035	JBUSH	*03:01		-	-	-	-
25	9049	IBW9	*02:02		-	-	-	-
26	9285	WT49	*02:01		-	-	-	-
27	9191	CH1007	*04:01	*05:01	-	-	+	-
28	9320	BEL5GB	*02:02	*03:01	-	-	-	-
29	9050	MOU	*02:02		-	-	-	-
30	9021	RSH	*04:02		-	-	+	-
31	9019	DUCAF	*02:01		-	-	-	-
32	9297	HAG	*03:01		-	-	-	-
33	9098	MT14B	*03:02		-	-	-	-
34	9104	DHIF	*03:01		-	-	-	-
35	9302	SSTO	*03:05		-	-	-	-
36	9024	KT17	*03:02		-	-	-	-
37	9065	HHKB	*06:03		-	-	-	-
38	9099	LZL	*03:01		-	-	-	-
39	9315	CML	*02:01	*03:01	-	-	-	-
40	9134	WHONP199	*02:02	*03:03	-	-	-	-
41	9055	H0301	*06:09		-	-	-	-
42	9066	TAB089	*06:01		-	-	-	-
43	9076	T7526	*03:03		-	-	-	-
44	9057	TEM	*05:03		-	-	-	-
45	9239	SHJO	*02:02		-	-	-	-
46	9013	SCHU	*06:02		-	-	-	-
47	9045	TUBO	*03:01		-	-	-	-
48	9303	TER-ND	*05:01		-	-	-	-



101.215-12 – including *Taq* pol., IFU-01
101.215-12u – without *Taq* pol., IFU-02

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Lot No.: 7H9

Lot-specific Information

¹The provided cell line HLA specificities are retrieved from the <http://www.ihwg.org/hla> web site. The specificity of an individual cell line may thus be subject to change.

²The specificity of each primer solution in the kit has been tested against 48 well characterized cell line DNAs and where applicable, additional cell line DNAs.

No DNAs carrying the alleles to be amplified by primer solutions 4 to 6, 9, 10, 12 to 18 and 20 were available. The specificities of the primers in primer solutions 4, 5, 9, 10 and 15 were tested by separately adding one additional 5'-primer, respectively one additional 3'-primer.

In primer solutions 6, 12, 14 and 16 it was only possible to test the 3'-primers, the 5'-primers were not possible to test. In primer solutions 13, 17, 18 and 20 it was only possible to test the 5'-primer, the 3'-primer was not possible to test. In primer solutions 3, 5, 9 and 11 one or more 5'-primers were not possible to test, and in primer solution 15 one 3'-primer was not possible to test. One additional 3'-primer in primer solution 11 was tested by separately adding one 5'-primer.

101.215-12 – including *Taq* pol., IFU-01
101.215-12u – without *Taq* pol., IFU-02

Visit <https://labproducts.caredx.com> for
“Instructions for Use” (IFU)

Lot No.: **7H9**

Lot-specific Information

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Fax: +46-8-717 88 18

E-mail: orders-se@caredx.com

Web page: <https://labproducts.caredx.com/>

CareDx Lab Solutions Inc., 901 S. Bolmar St., Suite R, West Chester, PA 19382

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